

PLUTONIUM UPTAKE BY THE GREEN ALGA Scenedesmus obliquus (TÜRP) KÜTZ AS A FUNCTION OF ISOTOPE AND OXIDATION STATE

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PLUTONIUM UPTAKE BY THE GREEN ALGA Scenedesmus obliquus (TÜRP) KÜTZ AS A FUNCTION OF ISOTOPE AND OXIDATION STATE

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## **ABSTRACT**

Uptake of <sup>238</sup>Pu<sup>4+</sup>, <sup>238</sup>Pu<sup>6+</sup>, <sup>239</sup>Pu<sup>4+</sup> and <sup>239</sup>Pu<sup>6+</sup> by the green alga *Scenedesmus obliquus* (Türp) Kütz was studied to determine whether isotope or oxidation state differences affect Pu uptake from aqueous medium by algal cells. At equivalent <sup>238</sup>Pu and <sup>239</sup>Pu concentrations, even when oxidation states differed, accumulations of these isotopes by *S. obliquus* were not significantly (p >0.05) different. Plutonium accumulation by *S. obliquus* was log-linear. Liquid scintillation detectors coupled to pulseheight analyzers were suitable for determining the alpha activity of samples and were accurate over a wide range of sample activities. Plutonium-237 was an effective tracer for determining plutonium chemical recovery.

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## INTRODUCTION

Though the separations chemistry of plutonium has been studied extensively, little is known about the environmental chemistry (24) of this potentially hazardous material. (32) Knowledge and understanding of the complex cycling of plutonium in the environment is necessary to predict the effects of possible releases. (7) Plutonium can exist as 15 isotopes with atomic masses between 232 and 246. At the present time, the isotopes of environmental importance are 238 Pu, 239 Pu, and 240 Pu. Plutonium can exist simultaneously in four valence states in solution (3+, 4+, 5+, and 6+). (23)

Recent studies have raised the question of differential availability of <sup>237</sup>Pu, <sup>238</sup>Pu, and <sup>239</sup>Pu to biological systems and of differential mobility in soil. <sup>(14,17,22)</sup> The difference in availability of plutonium isotopes may be attributed to several possible mechanisms. Oxidation state has been shown to be important in determining plutonium availability to plants, <sup>(24)</sup> and Bondietti et al. <sup>(8)</sup> suggest that knowledge of plutonium oxidation state may help to explain environmental findings.

Kinetic isotope effects are generally small, and depend on the relative magnitude of mass difference between the isotopes. (18) Kinetic isotope effects between <sup>237</sup>Pu, <sup>238</sup>Pu and <sup>239</sup>Pu are expected to be negligible. The major difference between the isotopes is in specific activity and decay energy. The specific activities (Ci/g) are <sup>237</sup>Pu, 12,080; <sup>238</sup>Pu, 17.2; and <sup>239</sup>Pu, 0.058. Plutonium-237 decays predominantly by electron capture (99%), while <sup>238</sup>Pu and

<sup>239</sup>Pu decay by alpha emission. The decay energies are 0.22 MeV for <sup>237</sup>Pu, 5.59 MeV for <sup>238</sup>Pu, and 5.24 MeV for <sup>239</sup>Pu. <sup>(19)</sup> These differences in decay properties can cause differential availability of the isotopes by at least three mechanisms

- Recoil: The greater decay energy of <sup>238</sup>Pu may result in increased ejection of atoms or clusters of atoms from PuO<sub>2</sub> particles or of atoms sorbed onto substrates such as soil. (15)

  The ejected atoms may be more mobile in the environment.
- Oxidation: The higher specific activity and greater decay energy of <sup>238</sup>Pu may provide opportunities for local oxidation to Pu<sup>6+</sup> by radiolysis products. <sup>(9)</sup> Pu<sup>6+</sup> is more available to plants <sup>(25)</sup> and is more mobile in soil. <sup>(26)</sup>
- Polymerization: The formation of polymeric hydrous oxides of plutonium is dependent on the plutonium concentration. (11,1)

  The plutonium concentration when 237Pu is used in experiments is usually much lower than it is when 238Pu or 239Pu is used.

  This may result in formation of plutonium polymer with 238Pu or 239Pu but not with 237Pu, resulting in increased availability of 237Pu. (6)

These mechanisms of differential availability of Pu isotopes will apply only when the isotopes are not homogeneously mixed. If the isotopes are present as a homogeneous mixture, the above mechanisms may result in increased plutonium availability but not in differential isotope availability.

This paper discusses an investigation of a simple interaction between a biological system and Pu. Specifically, an axenic algal culture, Scenedesmus obliquus, was exposed to the 4+ and 6+ oxidation states of  $^{238}\text{Pu}$  and  $^{239-240}\text{Pu}$  (herafter referred to as  $^{239}\text{Pu}$ ) at three Pu concentration levels, to determine the effects of oxidation state and isotope. Phytoplankton is one of the first trophic levels in which soluble compounds are involved, and algal species can quickly accumulate various trace elements including radionuclides deposited in aquatic systems. (13) Scenedesmus obliquus is a commonly occurring species in many fresh water systems, and is, therefore, of ecological importance. Wahlgren and Nelson<sup>(30)</sup> reported that sorption of plutonium by phytoplankton may be responsible for the seasonal loss of plutonium from the epilimnion of Lake Michigan. Additional objectives of this study were to develop a rapid and inexpensive method of plutonium determination in aquatic systems and investigate the use of 237Pu as a tracer for chemical yield determinations during sample preparation and analysis.

### METHODS AND MATERIALS

Algal Uptake

Axenic cultures of *Scenedesmus obliquus* (Türp) Kürtz (Strain No. 1592) were obtained from the Indiana University Culture Collection. (29) Cultures were checked for bacterial contamination by plating and microscopic examination. Stock cultures were grown in 4000-mL Erlenmeyer flasks containing 3500 mL of AAP culture

medium bubbled with sterile air.  $^{(2)}$  Cultures were maintained at 24 ±2°C in a controlled-environment chamber with 4,035-lux illumination from balanced-spectrum Growlux fluorescent bulbs on a 12 h of light - 12 h of dark regime. Cells were harvested from early stationary phase by continuous-flow centrifugation and resuspended in 500 mL of fresh AAP medium. Cells were centrifuged at 12062 RCF for 10 minutes over 28% by weight bis-2(ethylhexyl) sebacate in N-butyl phthalate to separate viable cells from dead cells and cell fragments. The recovered cells were resuspended in 350 mL of AAP medium, resulting in a stock with a density of 1 × 10 $^7$  cells/mL.

Plutonium-238 and  $^{239}$ Pu solutions were obtained from the Analytical Chemistry Division of the Savannah River Laboratory. The  $^{237}$ Pu solution was obtained from Oak Ridge National Laboratory. The  $^{237}$ Pu contained 1.8 nCi/ $\mu$ C  $^{237}$ Pu gross alpha activity. Results of analysis of the  $^{237}$ Pu are shown in Table 1. Plutonium solutions were prepared by dilution of stock  $^{238}$ Pu $^{4+}$ ,  $^{238}$ Pu $^{6+}$ ,  $^{239}$ Pu $^{4+}$ , and  $^{239}$ Pu $^{6+}$  solutions. The Pu $^{4+}$  stock solutions were prepared by evaporating a Pu solution to dryness and dissolving the residue in 10 mL of 1N HNO3. The solution was heated to  $70^{\circ}$ C in a water bath and 0.5 mL of 1M NaNO2 was added. The solution was maintained at  $70^{\circ}$ C for one hour and then diluted to 25 mL with 1N HNO3. Pu $^{6+}$  was prepared by adding 0.05 mL of 0.2M KMnO4 to 25 mL of a Pu solution (in 1N HNO3). The solution was allowed to stand for two hours and then 0.3M MnCl2 was added dropwise until

MnO<sub>2</sub> formed. The solution was allowed to stand for an additional two hours and was then filtered through a 0.45- $\mu$ m filter. The purities of the oxidation states were checked by extraction into thenoyl trifluoroacetone (TTA) from 1N HNO<sub>3</sub>. Under these conditions, only Pu<sup>4+</sup> will extract. (11) TTA-xylene extracted >97% of the plutonium from the Pu<sup>4+</sup> solutions and <0.1% from the Pu<sup>6+</sup> solutions.

Plutonium sorption by algal cells was studied in silicone-coated glass, 250-mL Erlenmeyer flasks containing 93 mL of sterile AAP medium. One mL of plutonium solution in 1N HNO $_3$  was pipetted into each flask and the pH was adjusted to 5.5 with 1N NaOH. Five mL of algal suspension were added, resulting in a total volume of 100 mL and an algal cell density of approximately 5.5  $\times$  10 $^5$  cells/mL. The ionic strength (I) of the culture medium was 0.0143. Using the Davies approximation, the resulting activity coefficients for Pu $^{4+}$ , Pu $^{0}$ <sub>2</sub>, Pu $^{0}$ <sub>2</sub>OH $^{+}$ , and Pu $^{0}$ <sub>2</sub>(OH) $^{-}$ 3 were 0.27, 0.72, 0.92, and 0.92, respectively.

After a 4-h incubation, two 5-mL aliquots were taken for microscopic cell density determination and plating loss. One 10-mL aliquot was removed from control flasks to determine plating loss in the absence of algal cells. The remaining algal suspension was transferred, in 30-mL aliquots, to a polyethylene centrifuge tube and centrifuged at 12062 RCF for 15 minutes over 28% by weight bis-2-(ethylhexyl) sebacate in N-butyl phthalate. (12) The supernate was aspirated off and retained. The algal cells and bis-2-(ethylhexyl) sebacate were washed from the centrifuge tube with

acetone and retained separately.

A  $^{239}\text{Pu}$  concentration comparable to the lowest  $^{238}\text{Pu}$  concentration is approximately 0.4 d/m/mL which is below detectability of the counting apparatus. 239Pu, therefore, did not overlap at this level. No attempt was made to achieve a 238Pu concentration similar to the highest 239Pu level, because that high activity would present counting problems and safety hazards. In addition, there was one control blank per treatment combination containing no algae. The experimental units were arranged in a 2  $\times$  2  $\times$  3 design with 5 replicates of each treatment combination (Table 2). Thus, the experiment consisted of 60 experimental units and 12 control blanks. However, because the centrifuge only held eight samples concurrently, samples were randomly blocked orthogonally into groups of eight and spaced  $1-1/2\ h$  apart for incubation and algal cell harvesting. Handling of controls was similar to that of experimentals, to evaluate possible contamination during the separation procedure.

# Analytical Procedures

Radiochemical analysis involved a combination of wet and dry ashing (Boni, A. L. personal communication, 1976). The <sup>237</sup>Pu stock solution (Table 1) was diluted 1000-fold, and one-mL aliquots were added to each sample before ashing. Samples were evaporated to dryness in 10 mL of 8N HNO<sub>3</sub>. Following dry ashing at 500°C in a muffle furnace for six hours, samples were twice dissolved in 15 mL of 8N HNO<sub>3</sub> and taken to dryness. The final residue was dissolved in 3 mL of hot 1N HNO<sub>3</sub> and a 2-mL aliquot was pipetted into

a liquid scintillation vial containing 20 mL of Aquasol-2 scintillation solution (New England Nuclear). Samples were counted for 10 to 1000 minutes, depending upon sample activity.

Two liquid scintillation systems were used for plutonium measurement by alpha counting: 1) Nuclear Chicago liquid scintillation counter interfaced to an Ortec multichannel analyzer; and 2)
Packard Tri-Carb liquid scintillation counter interfaced to a
Technical Measurements Company multichannel analyzer. The liquid scintillation systems had backgrounds of 23 cpm with 2 SE minimum detectability of 0.5 cpm. Although these systems did not have the excellent pulse resolution characteristics of custom experimental systems, (20) they detected low-activity 238Pu and 239Pu samples quite effectively. 237Pu tracer concentration in each sample was determined using a 20-cm NaI well-type detector.
Taking the small 237Pu alpha contribution (Table 1) into account, the total sample plutonium was calculated using Equation 1.

Total sample plutonium (d/m) = 
$$\left[ sa^{Pu}_{\alpha} - \left( \frac{st^{237}Pu}{st^{9}u}_{\alpha} \right) \left( \frac{237}{st^{9}u}_{\gamma} \right) \right] \left( \frac{237}{st^{9}u}_{\gamma} \right)$$
(1)

where:

 $_{\text{sa}}^{\text{Pu}}{}_{\alpha}$  and  $_{\text{st}}^{\text{237}}\text{Pu}_{\alpha}^{\text{}}$  = net alpha counts of sample and

<sup>237</sup>Pu standard in counts/min

 $_{sa}^{Pu}{}_{\gamma}$  and  $_{st}^{2\,3\,7}{}^{Pu}{}_{\gamma}$  = net gamma counts of the sample and  $_{st}^{2\,3\,7}{}^{Pu}$  standard in counts/min

Data were converted from alpha decays/min per sample to plutonium atoms using Equation 2.

Pu atoms = 
$$\frac{d/m}{SW(2.22 \times 10^{12}) d/m/Ci} \cdot A$$
 (2)

where:

d/m = alpha activity of sample (decays/min)

S = specific activity of isotope (Ci/g)

W = atomic mass of isotope (g/gram atom)

A = Avogadro's number  $(6.023 \times 10^{23} \text{ atoms/gram atom})$ Plutonium in solution is reported as plutonium atoms/mL and plutonium associated with algal cells is reported as plutonium atoms/cell which facilitates direct comparison of  $^{238}$ Pu and  $^{239}$ Pu. Thus, the "pCi/g" term was eliminated as it presented a distorted representation of isotopic behavior when comparing isotopes of widely differing half-lives, such as  $^{238}$ Pu (88 yr) and  $^{239}$ Pu (24,000 yr).

### **RESULTS**

The experimental results of plutonium uptake by the green alga  $S.\ obliquus$  are summarized in Figure 1. The amount of  $^{238}Pu^{4+}$  and  $^{238}Pu^{6+}$  associated with algal cells was log-linearly proportional to Pu concentration in the medium. No significant (p >0.05) oxidation state effect on algal sorption of  $^{238}Pu$  or  $^{239}Pu$  was observed at the three concentrations studied. Likewise, no significant (p >0.05) difference between the algal uptake of  $^{238}Pu$  and  $^{239}Pu$  was observed at similar concentrations. Approximately 20 percent of the plutonium was observed associated with the algal fraction.

Plutonium is also known to "plate out" on vessel walls and be

effectively removed from solution. However, when each entire solution and its associated algal fraction were assayed and taken together, a 96.5% mass balance was obtained, indicating a rather small loss due to adsorption on vessel walls.

### DISCUSSION

The experimental plutonium concentrations are relevant to levels encountered in the environment. The lowest  $^{238}$ Pu level studied ( $10^8$  atoms/mL) is just above plutonium concentrations reported in the Savannah River (approximately  $10^7$  atoms/mL). (3) The maximum permissible  $^{239}$ Pu concentration in drinking water ( $^{16}$ ) is  $10^9$  atoms/mL.

Plutonium can coexist in solution at all four oxidation states  $^{(4)}$  between Pu<sup>4+</sup> and Pu<sup>6+</sup>. The distribution of oxidation states at equilibrium is dependent on the pH and Eh of the solution and on the degree of complexation.  $^{(2\,8)}$  The estimated oxidation state distribution in the AAP medium (pH = 5.5, Eh = 0.35v, 0.01M NO $_3$ ) is 100% Pu<sup>3+</sup>. However, the rate at which equilibrium is attained will be different depending on the initial oxidation state. No analyses were conducted subsequent to the experiments to verify that the oxidation state of the isotopes had remained constant throughout the 4-h experiment. Quadrivalent plutonium will be reduced to Pu<sup>3+</sup> more readily than will  $(\text{PuO}_2)^{2^+}$  because two plutonium-oxygen bonds must be broken in the latter case.  $^{(11)}$  Therefore, it is unlikely that both Pu<sup>4+</sup> and  $(\text{PuO}_2)^{2^+}$  converted in exactly the same manner during the four-hour incubation.

Alberts (personal communication, 1977\*) found  ${\rm Pu}^{6}{}^+$  To be the most stable oxidation state in filtered and synthetic Lake Michigan water.

Plutonium ions will form complexes with nitrate ions in solution. The degree of complexation is dependent on the oxidation state of the Pu and NO<sub>3</sub> concentration. (21) The order of stability of nitrate complexes (11) is Pu<sup>4+</sup>>Pu<sup>3+</sup>>(PuO<sub>2</sub>)<sup>2+</sup>>(PuO<sub>2</sub>)<sup>+</sup>. While Pu<sup>4+</sup> readily forms nitrate complexes, nitrate exhibits only a slight tendency (11) to form complexes with Pu<sup>6+</sup>. The degree of complexation in the AAP medium is difficult to estimate because of the low ionic strength and low NO<sub>3</sub> concentration, since reported Pu stability constants were measured in solutions of ionic strength 1M or greater and at relatively high NO<sub>3</sub> concentrations. (11) If, due to kinetic considerations, equilibrium had not been established during the 4-h incubation, the solubility, extent of hydrolysis and degree of nitrate complexation of plutonium in the two solutions (initially Pu<sup>4+</sup> and Pu<sup>6+</sup>) could be considerably different. (27)

Plutonium is known to polymerize in weakly acidic solutions resulting in polymers<sup>(11)</sup> of Pu<sup>4+</sup> which can be removed by centrifugation at 3600 rpm for 2.5 h. However, polymerization is dependent upon Pu concentration as well as pH. If only small quantities of Pu are available, a high pH is required to induce polymerization. Thus, polymerization was probably not an important factor at the Pu concentrations studied here.

The results of this study show that from a practical standpoint of predicting environmental behavior in a water-phytoplankton system, whether the Pu<sup>4+</sup> and Pu<sup>6+</sup> solutions reached equilibrium (resulting in the same distribution of oxidation states) or not is immaterial. In either case, whether uptake is independent of oxidation state or equilibrium is reached in as little as four hours, the result, Pu absorption by algae, would be the same. The distribution coefficients of Pu<sup>6+</sup> and Pu<sup>4+</sup> in seston and sediments are such that they suggest that Pu<sup>6+</sup> may be reduced to Pu<sup>4+</sup> by algal cells at the cell wall (Alberts, personal communication, 1977). Such a mechanism could also explain the results observed here. For determining and modeling plutonium uptake by algae, a knowledge of the  $(CO_3^{2-})$  is necessary for determining plutonium availability to phytoplankton. (10)

Bair et al. <sup>(6)</sup> found differential mobility of <sup>237</sup>Pu and <sup>239</sup>Pu in dogs. Bair <sup>(5)</sup> also found <sup>238</sup>PuO<sub>2</sub> translocation in dogs to be greater than <sup>239</sup>PuO<sub>2</sub>. While this biological system is much different than that studied here, it does indicate differential behavior due to mass differences. However, Wayman and Bartelt <sup>(31)</sup> reported no significant difference in <sup>238</sup>Pu and <sup>239</sup>Pu concentration factors in fish, when expressed on a gram atomic mass basis. Their results agree closely with those of this study, which indicates no effect of Pu mass on sorption by algal cells.

The sample preparation and counting methods were simple, inexpensive, and suitable for this experiment.  $^{2\,37}$ Pu, as the internal spike, eliminated the need for sophisticated counting systems, such as high-resolution alpha spectroscopy systems that are required when  $^{2\,36}$ Pu or  $^{2\,42}$ Pu are used as the internal spike. The technique has applicability to a variety of aquatic experimental designs where the gross alpha content is solely from plutonium added by the experimenter.

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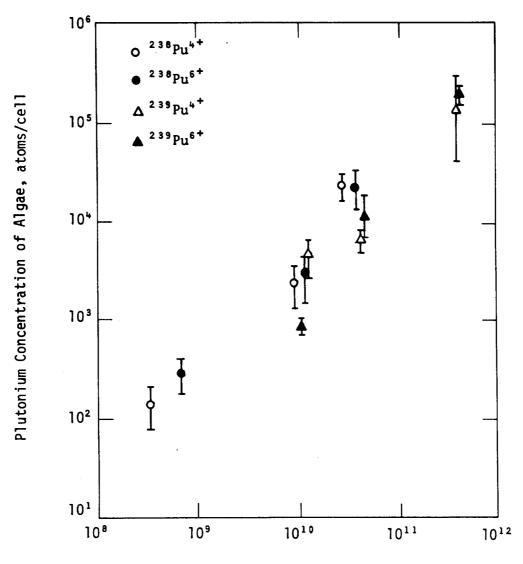
Table 1. Analysis of Plutonium-237 Product as of November 4, 1976

<sup>237</sup> Pu	1.4 µCi/ml
Solution	0.5 M HC1
<sup>67</sup> Ga (78 hr)	0.8 nCi/µCi <sup>237</sup> Pu
Gross Alpha Activity	1.8 nCi/μCi <sup>237</sup> Pu
Alpha-Spectrum Components	4.9 MeV ∿9.2%
	5.3 MeV ∿13.7%
	5.7 MeV ∿71.1%

Table 2. Plutonium Nitrate Concentrations in Experimental Flasks $^{lpha}$ 

Isotope and Oxidation State	Pu Concentration, atoms/mL
<sup>2 3 8</sup> Pu <sup>4 +</sup>	$3.25 \times 10^{8} \text{ (n=5); } 9.15 \times 10^{9} \text{ (n=5); } 2.71 \times 10^{10} \text{ (n=5)}$
<sup>238</sup> Pu <sup>6+</sup>	$6.87 \times 10^{8} \text{ (n=5); } 1.11 \times 10^{10} \text{ (n=5); } 3.78 \times 10^{10} \text{ (n=5)}$
<sup>2 3 9</sup> Pu <sup>4 +</sup>	$1.11 \times 10^{10} (n=4); 3.87 \times 10^{10} (n=5); 3.78 \times 10^{11} (n=3)$
<sup>2 3 9</sup> Pu <sup>6 +</sup>	$1.04 \times 10^{10} (n=3); 4.61 \times 10^{10} (n=5); 3.81 \times 10^{11} (n=5)$

 $<sup>\</sup>alpha$ . n = number of replicate flasks.



Plutonium Concentration in Water, atoms/mL

FIGURE 1. Plutonium Uptake by Green Alga S. obliquus